



VERIFICATION OF TRANSLATION

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Method for the production of an immunostimulating mucin (MUC1)

The present invention relates to a method for producing or identifying a MUC1 molecule that is able to generate an immune response in humans. The invention also relates to a method for producing or identifying a cell, cell lines, or cell lysates comprising a MUC1 molecule that is able to generate an immune response in humans. The invention further relates to methods for producing medicaments and diagnostic agents. A further object of the present invention is the use of the MUC1 molecules, cells or cell lysates obtainable by means of the methods of the present invention for producing a medicament used for the treatment or prevention of tumours. The invention also relates to a purified MUC1 molecule that can be obtained by means of the methods of the present invention and has an immunostimulating effect on humans. The invention additionally relates to the use of an MUC1 antibody for the production of a medicament used for the treatment or prevention of tumours.

In the past, the human mucin MUC1 was inter alia also called polymorphic epithelial mucin (PEM). At the moment it is the only mucin of which it is known that it is a transmembrane glycoprotein. The polypeptide of the MUC1 consists extracellularly mainly of repetitive units, the so-called tandem repeats, their number in the human population being polymorph for genetic reasons. In agreement with the general characteristic of the mucins, MUC1 also carries a large number of carbohydrate chains joined by O-glycosidic bond. Each tandem repeat consists of 20 conserved amino acids (NH₂-HGVTSAPDTRPAPGSTAPPA-COOH) with 5 potential O-glycosylation sites (two serines and three threonines). Until recently the general state of knowledge was that the potential glycosylation site in the PDTR sequence is non-glycosylated (e.g.: Finn OJ, Jerome KR, Henderson RA, Pecher G, Domenech N, Magarian-Blander J, Barratt-Boyes SM. MUC-1 epithelial tumour mucin-based immunity and cancer vaccines. Imm Revs 145:61-89 (1995), p. 67).

One of the reasons given for this was that after immunization of mice with MUC1 from tumour or non-tumour sources almost exclusively antibodies were generated which are directed against the PDTR region, which therefore is also called immunodominant region. A further reason was that at this time in-vitro glycosylation attempts were not successful in inducing glycosylation in the PDTR region. Recently, however, it was shown in particular by mass spectrometric analysis of MUC1 from human milk (non-tumour source) that the threonine of the PDTR region of some tandem repeats can be in glycosylated form although others are present in an unglycosylated form. Furthermore, the analysis of a group of MUC1 specific antibodies has shown that a glycosylation in the PDTR region enhances or even first makes possible the binding of individual antibodies Ab to short MUC1 peptides vis-à-vis the unglycosylated peptide, whereas others bind to this site completely independently of glycosylation.

MUC1 per se is an established tumour marker. The following known properties characterize MUC1 as a tumour marker: In the normal epithelium, especially in the epithelium of glands, MUC1 is expressed exclusively polar on the apical membrane of the cells. However, this strict apical localization is lost during cancerogenesis, and it comes to an apolar expression over the whole cell surface, whereby the MUC1 becomes accessible to the immune system. It is a further characteristic of MUC1 that its expression is increased during cancerogenesis. As third known tumour property the glycosylation of MUC1 changes during cancerogenesis resulting in an aberrant glycosylation. This results on the one hand in the expression of de novo carbohydrate antigens and on the other hand in the partial exposure of peptide epitopes (Karsten, Cancer Research 58, 2541-2549, 1998; Price, Tumor Biol. Suppl1: 1-5, 1998). As described in the prior art, MUC1 is a molecule known per se which is well-established as a tumour marker. However, MUC1 is not a strictly structurally defined molecule but a heterogeneous mixture of molecules. The heterogeneity is caused by various properties of MUC1:

- a. MUC1 is genetically polymorph in the human population as regards the number of tandem repeats.
 - b. Glycosylation and expression vary in different tissues and at different differentiation stages.
 - c. MUC1 preparations from tissue or cell lines show a high degree of heterogeneity of carbohydrate structures and thus of MUC1 molecules.
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- d. MUC1 preparations from tissue or cell lines show that within one MUC1 molecule the tandem repeats are heterogeneously glycosylated, this applies to the sugar chains as well as to the glycosylation sites.
- e. The techniques available at present do not allow the correlation of a carbohydrate chain to a specific tandem repeat or to a specific glycosylation site of MUC1 from native material.
- f. So far glycosylation has not been completely described for any MUC1.

The chemical structure of a natural MUC1 molecule is in no case completely known. To the contrary, the glycosylation is only known in few cases and there only partially. This shows that the conventional native MUC1 is no defined structure but a large heterogeneous group of molecules.

In addition, in case of cancer by changes in the glycosylation apparatus changes occur in the MUC1 glycosylation which introduce a further complexity into the heterogeneity. This changed glycosylation is again strongly heterogeneous in relation to the type of tumour, within one tumour type, with regard to carbohydrate chains, and to the distribution of the carbohydrate chains.

It has been described that tumour-associated MUC1 acts immunosuppressive on T-cell proliferation (Agrawal et al., Nat. Med., 1998, Oct. 4(10):1093). It has been further described that this relates especially to longer MUC1 molecules having more than 60 amino acids, which were obtained from tumour cells (WO 99/23114). A corresponding immunosuppression is particularly disadvantageous or prevents the use of corresponding tumour-associated MUC1 molecules expressed from cells. In particular it has been described that MUC1 obtained by means of the antibody B27.29 suppresses the immune system (US20020044943, WO 9810783). For a suitable and effective tumour therapy or prophylaxis on the basis of the tumour marker MUC1 it is necessary to find a way to produce MUC1 molecules which do not suppress the immune system at all or only to a limited extent, and which, preferably, even act as stimulants to the immune system. An established tumour therapy or prophylaxis on the basis of the tumour marker MUC1 is not known.

The technical problem underlying the present invention is thus to provide measures for therapy and/or prophylaxis of tumours associated with the tumour marker MUC-1.

This technical problem is solved by the embodiments described in the claims.

The present invention thus relates to a method for producing an MUC1 molecule which is able to generate an immune response in humans, comprising:

- (a) contacting a mixture of MUC1 molecules with an antibody having the following properties:
 - i) binding to the immunodominant region of the MUC1 tandem repeat; and
 - ii) the binding to an MUC1 fragment having a length of between 9 and 40 amino acids and containing sequences of the MUC1 tandem repeat and the immunodominant region is made possible or enhanced by glycosylation of the threonine of a PDTR sequence; and
 - iii) the binding of unglycosylated MUC1 tandem repeats is enhanced if the unglycosylated MUC1 tandem repeats exist in series; and
 - iv) the binding to multiple glycosylated MUC1 tandem repeats carrying glycosylations at the threonines of several PDTR sequences of the immunodominant region is enhanced vis-à-vis the short MUC1 fragments of ii) in view of an additive effect of length and the PDTR glycosylation;for a period of time that is sufficient and under conditions that are suitable to form an immune complex;
- (b) isolation of the immune complex; and
- (c) providing an MUC1 molecule from the immune complex.

In addition to the previously explicitly mentioned steps the term "preparation" also includes additional steps, such as pretreatment of the starting material or further treatment of the final product. Pretreatment procedures will be described in more detail in the following, further treatment procedures comprise e.g. the final formulation of the prepared MUC1 molecule in suitable dosage forms. The kind of dosage form, e.g. tablet, solution or lyophilisate, depends on the intended use. The person skilled in the art knows which dosage form is suitable for which application. Depending on the dosage form used the MUC1 molecule produced by the method of the invention may be present together with excipients, carrier substances or further active agents. Here, excipients are preferably adjuvants, further active

agents are preferably immunostimulating molecules, such as interleukins. The MUC1 molecule produced by a method according to the present invention may also be chemically modified in further treatment steps. Here, the MUC1 molecule is preferably combined with one or more further molecules in a suitable manner, i.e. by chemical or physical interaction. Preferably serving as further molecules within the meaning of the invention are other proteins or peptides which are fused with the MUC1 molecule prepared according to the method of the invention. In this case the MUC1 molecule is present as fusion protein. Further molecules within the framework of the inventive method are in particular substances having a therapeutic or diagnostic activity, such as radioisotopes or toxins. Further molecules to be used in connection with the invention are such substances which are suitable to serve as carrier materials or which may serve as components to couple the MUC1 molecules with carrier substances. Such carrier materials or coupling components are known from the prior art.

The term "MUC1 molecule which is able to generate an immune response in humans" comprises MUC1 molecules or fragments thereof which belong to a species of MUC1 glycoproteins which alone or in combination with suitable adjuvants and/or immunostimulating molecules cause an immune response to MUC1-positive tumour cells. An immune response within the meaning of the present invention is the triggering of a humoral immune response and/or cellular immune response. A humoral immune response in the sense of the present invention means the inducing of antibodies by MUC1 molecules produced according to the method of the invention. A cellular immune response means the inducing of immune cells, e.g. CD4-positive and/or CD8-positive cells, by the MUC1 molecules produced according to the method of the present invention, which directly or indirectly fight MUC1-positive tumour cells. The MUC1 molecules are characterized by the fact that they have no or only a weak immunosuppressive effect on T-cells. Preferably used are MUC1 molecules which stimulate T-cells, i.e. these molecules have a stimulating effect within the meaning of the present invention. The MUC1 molecules are preferably tumour-associated. The immunostimulating properties of MUC1 molecules and induced immune responses can be shown in-vitro and in-vivo by at least one of the methods described in the Examples or other methods known to the skilled person, such as ELISA, ADCC- and CDC-assays, T cell stimulation assays and NOD-SCID mouse models. The skilled person is therefore readily able to determine whether certain MUC1 molecules belong to the same species with respect to their immunostimulating properties. Preferably the members of this

species share in one or more tandem repeat(s) chemical properties which are defined in the following in more detail:

The term “contacting” comprises all measures necessary to allow a chemical, biochemical or physical interaction between the MUC1 molecule and the antibodies used in the method. For this purpose the MUC1 molecules and the antibodies are preferably introduced into a suitable liquid. Suitable liquids are e.g. buffer solutions which, as described in the prior art or the Examples, in ELISA or RIA detection systems or solutions described for chromatographic purification methods. Particularly suitable are solutions used in the prior art for affinity chromatography with antibodies. Suitable solution are particularly preferably the solutions described in detail in the Examples.

The term “mixture of MUC1 molecules” comprises a large number of MUC1 molecules which can belong to the same or different species. As already stated before the MUC1 tumour markers described in the prior art are a mixture of different molecular species which differ as to their glycosylation and partly in their amino acid composition. The mixture of MUC1 molecules used in the inventive method may also contain further molecules, such as sugars, lipids, proteins or peptides which do not belong to the MUC1 group. Such mixtures of MUC1 molecules can preferably be obtained from tumour tissue, tumour cells as well as from cells or cell lines producing inter alia tumour-associated MUC1. In a particularly preferred manner lysates may be produced from these tissues, cells or cell lines. These cell lysates or fragments thereof may also serve as MUC1 mixtures within the meaning of the present invention. Synthetic glycopeptides or fragments of the tumour-associated MUC1 may also serve as mixture of MUC1 molecules. The preparation of synthetic glycopeptides is known from the prior art (Karsten, loc. cit.). Glycopeptides may also be produced by treatment with proteases and/or glycosidases from MUC1 preparations.

The term “antibody” within the meaning of the invention comprises murine, chimeric, humanized, human antibodies, whole antibodies of different isotypes or fragments of antibodies, such as single-chain antibody fragments (ScFv), Fab fragments, multivalent antibody fragments (e.g. diabodies, triabodies or tetrabodies), or antibody fragments of other formats, even such as are used as fusion proteins coupled with other peptides or protein structures, as well as antibodies or antibody

fragments which, besides the specificity to MUC1 according to the present invention also have one or more further specificities, such as bispecific antibodies. The specificity according to the present invention of the antibodies according to the present invention is defined further below. These antibodies are produced by techniques known per se. The antibodies used in the method according to the present invention are characterized by the above mentioned properties.

In this connection the term "immunodominant region of the MUC1 tandem repeat" within the meaning of the present invention means the PDTR region which strictly speaking also comprises further amino acids of the adjacent sequences of the MUC1 tandem repeats, for example APDTRPAP. Whether an antibody binds to this region can be readily determined by the skilled person by methods known from the prior art.

The term "MUC1 tandem repeat" designates a sequence of 20 amino acids: HGVTSAPDTRPAPGSTAPPA. The five potential O-glycosylation sites on two serines and three threonines have been underlined in the previously shown sequence. Furthermore, in accordance with the present invention this term also comprises such variants which possess mutations and thus amino acid substitutions in one or more tandem repeats of a longer MUC1 molecule. The tandem repeat has 20 amino acids and may start in any place.

The term "unglycosylated MUC1 tandem repeats exist in series" designates a linear sequence of several tandem repeats of the MUC1, this sequence not being interrupted by other sequences. Preferably, there exist at least two, at least three, at least four or at least five tandem repeats existing in series. Most preferably there exist at least six tandem repeats in series.

The term "enhanced" within the meaning of the present invention means a binding of the antibody to a PDTR-glycosylated MUC1 peptide having between 9 and 40 amino acids that is significantly stronger than the binding to a corresponding peptide which is unglycosylated in the PDTR region. This applies for PDTR-glycosylated tandem repeats under the above item ii (ii – binding to an MUC1 fragment having a length of between 9 and 40 amino acids and containing sequences of the MUC1 tandem repeat and the immunodominant region is enabled or enhanced by glycosylation of the threonine of a PDTR sequence). This also includes a binding of the antibody to several tandem repeats in series which is significantly stronger than a binding to individual tandem repeats. This applies to unglycosylated tandem

repeats under item iii) (binding to unglycosylated MUC1 tandem repeats is enhanced when the unglycosylated MUC1 tandem repeats exist in series;) and to PDTR-glycosylated tandem repeats under item iv) (iv) – the binding to multiple glycosylated MUC1 tandem repeats carrying glycosylations at the threonines of several PDTR sequences of the immunodominant region is enhanced vis-à-vis the short MUC1 fragments of ii) in view of an additive effect from their length and the PDTR glycosylation). Whether a binding is significantly enhanced may easily be determined by the skilled practitioner by suitable statistical tests and is shown in Examples 1 to 3.

The term “made possible” within the meaning of the present invention stands for the binding of an antibody to a PDTR-glycosylated MUC1 peptide having between 9 and 40 amino acids which is made possible by glycosylation at this site. Corresponding peptides which are not glycosylated at the PDTR region do not bind or the binding is hardly detectable with the methods used (see also 2).

The term “immune complex” within the meaning of the present invention means the product of an antibody-antigen reaction. Hereby the antibody reversibly binds to the antigen via physico-chemical interactions. The interacting components form the immune complex.

The term “isolation” comprises all measures or methods which serve to separate the immune complex from antibodies which have not formed an immune complex with an antigen as well as from MUC1 molecules or other molecules which were not bound by an antibody. These methods are known in the art and described in the Examples. They comprise for example measures leading to the precipitation of the immune complex or chromatographic methods. Further methods are also described in more detail in the following.

In addition the invention describes a novel and unexpected tumour epitope on the whole molecule of MUC1 which can be prepared/obtained by the method of the invention. The novel MUC1 tumour epitope according to the present invention on the whole molecule of MUC1 is characterized by the combined binding specificities of MUC1-specific monoclonal antibodies of group 1 which in the following are described in detail. All analyzed monoclonal antibodies having the common binding properties described in the following, were generated by immunization of mice with MUC1 from tumour material.

Antibodies of group 1 have the common relevant binding properties 1 to 4:

- 1) The antibodies bind to the immunodominant region (so-called PDTR region – see definition) of the MUC1 tandem repeat.
- 2) The binding of the antibodies to an MUC1 fragment of the MUC1 tandem repeats containing the PDTR region and having a length between 9 and 40 amino acids is enabled or enhanced by glycosylation on the threonine of a PDTR sequence of the immuno-dominant region. The binding to the corresponding MUC1 peptides which are not glycosylated in the PDTR sequence is clearly lower. A glycosylation with Tn (GalNAc α 1-O-Thr) is sufficient to achieve this effect, however a longer glycosylation as for example by TF (Gal β 1-3GalNAc α 1-O-Thr) is also possible to achieve this effect. Glycosylations on other glycosylation sites of the MUC1 tandem repeat have no or no significant influence on the basic effect (see Example 1 and Table 1).
- 3) The antibodies bind to unglycosylated MUC1 tandem repeats better if they exist in series. There the multiple unglycosylated MUC1 tandem repeat peptide comprises at least 3 tandem repeats, it being, however, also possible that the binding can only be strengthened by further tandem repeats and/or be strengthened even more significantly by an increased number of tandem repeats. With most antibodies the greatest increase in binding is found with MUC1 peptides having 4 tandem repeats to MUC1 peptides having 6 tandem repeats (see Example 2 and Table 2).
- 4) The antibodies show a binding property which in the invention is called an additive effect. The antibodies bind better to PDTR-glycosylated MUC1 tandem repeats when they exist in series. There the multiple PDTR-glycosylated MUC1 tandem repeat peptide contains at least 2 tandem repeats and at least 2 glycosylated PDTR regions. It is also possible that the binding is only strengthened by further PDTR-glycosylated tandem repeats and/or even more significantly increased by an increased number of PDTR-glycosylated tandem repeats. With many antibodies the greatest increase of binding is found with multiple PDTR-glycosylated MUC1 tandem repeat peptides of 4 tandem repeats to such of 6 tandem repeats (see Example 3 and Table 3).

The tumour epitope is described by the binding specificities of the antibodies which have the binding properties 1 and 2 and 3 and 4. Here preferred is the antigenic epitope structure which is bound by antibodies with particularly high additive effect, such as A76-A/C7 and VU-11E2, with A76-A/C7 being preferred. A further preferred

form of the MUC1 tumour epitope according to the invention is the structure which is preferably bound by the antibodies having a particularly high additive binding effect, such as A76-A/C7, VU-11E2, with A76-A/C7 being preferred, since in colon carcinoma patients A76-A/C7 shows a significantly reduced binding to MUC1 released in serum (shedded MUC1). In contrast, HMFG-1 shows a significantly stronger binding in colon and pancreas carcinoma patients. This fact is a great advantage for immune therapies using MUC1 molecules or MUC1 specific antibodies with this specificity.

Since the effect of the length of antibodies vis-à-vis unglycosylated tandem repeats alone does not bring about a difference between antibodies prepared by immunization with tumour material and such antibodies prepared by immunization with non-tumour material, it becomes clear that the loop structure described in the literature which occurs in unglycosylated MUC1 peptides of multiple tandem repeats in the form described therein does not exactly correspond to the tumour epitope according to the present invention.

All tested antibodies from immunizations with MUC1 from tumour material show a binding property which is dependent on the glycosylation of the PDTR region, preferably with an MUC1 structure having a length of at least 9 amino acids and being surrounded by at least 3 additional amino acids around the PDTR region, and on that unglycosylated MUC1 tandem repeats exist in series. The majority shows the additive binding effect from multiple tandem repeats and PDTR glycosylation. In contrast, nearly all tested antibodies obtained by immunization with non-tumour material are independent on glycosylation. Most antibodies are negative for the additive effect.

Preferred antibodies for the definition according to the invention and description of the tumour epitope according to the invention and the use according to the invention of these antibodies for the diagnosis and treatment of tumours as well as to the preparation according to the invention of suitable MUC1 molecules for the treatment and diagnosis of tumour, are such antibodies from group 1 which have an additive binding effect from length and PDTR glycosylation, as described under binding property 4, for example A76-A/C7, VU-11E2, VU-11D1, BC4E549, VU-12E1, VU-3D1 and b-12 (see Table 3; source of antibodies is disclosed in Table 1). These antibodies have an increased affinity to multiple tandem repeats and multiple PDTR glycosylated tandem repeats and to native tumour-MUC1 consisting of multiple tandem repeats, and are thus to be preferred according to the invention.

Particularly preferred are antibodies for the inventive definition and description of the tumour epitope and for the use according to the invention of these antibodies for the diagnosis and treatment of tumours as well as to the preparation according to the invention of suitable MUC1 molecules for the treatment and diagnosis of tumours, which have a pronounced additive binding effect. These antibodies bind to multiple PDTR glycosylated MUC1 tandem repeat peptides of 4) clearly to many times stronger than to short PDTR glycosylated MUC1 peptides of 2) and their binding to multiple unglycosylated MUC1 tandem repeats of 3). The antibodies of this preferred subgroup show a pronounced additive effect of binding enhancement by glycosylation of the PDTR sequence according to 2) and multimerisation of the tandem repeats according to 3) (see Example 3 and Table 3). The antibodies of this group, preferably A76-A/C7, VU-11E2 and VU-11D1 (see Example 3 and Table 3), show a particularly strong additive effect and have a particularly increased affinity to multiple PDTR glycosylated tandem repeats and thus to native tumour MUC1 which consists of multiple tandem repeats, and thus are preferred according to the present invention.

All tested MUC1-specific antibodies which have binding properties of group 1 correlate to the immunogen which was used for their generation in mice (see Example 2 and Table 1), for example in form of whole tumour cells or MUC1 obtained from tumour material. MUC1-specific monoclonal antibodies (mab) which were obtained from immunizations with non-tumour relevant MUC1 material, do not show this combination of binding properties. They are all independent or inversely (negatively) dependent on the glycosylation of the PDTR region according to 2) or independent from the length of the unglycosylated MUC1 peptides and thus the number of tandem repeats according to 3). This unexpected correlation shows that antibodies possessing the binding properties of group 1 recognize a specific antigenic structure on the MUC1 which corresponds to a novel tumour epitope on MUC1 and which is therefore particularly suitable as target structure for a tumour therapy and for diagnostic purposes. Therapeutic and diagnostic methods using this novel and unexpected tumour epitope as target structure, significantly improve previous approaches looking at the MUC1 according to its previously known tumour-associated properties overexpression, apolar expression and aberrant glycosylation.

The present invention describes a novel and surprising tumour epitope on the human polymorphic epithelial mucin MUC1. The invention claims this epitope as well as substances and methods for their preparation, which contain this epitope,

such as cells, glycoproteins or glycopeptides, and methods for use in the prophylaxis and treatment of tumour diseases, including vaccines, such as whole cell vaccines, glycoprotein vaccines or glycopeptide vaccines, as well as antibodies obtained with the assistance of MUC1 as immunogen obtained by the production method according to the present invention, and which have an increased additive effect and are therefore particularly suitable for use in the treatment and/or diagnosis of tumour diseases.

For use of MUC1 in tumour therapy and prophylaxis it is necessary to isolate and/or identify such MUC1 molecules which bear tumour-relevant epitopes and have as far as possible no or at least a reduced immuno-suppressive influence. This also applies to the isolation and preparation of cells or other carrier substances which carry MUC1 molecules that are suitable according to the present invention. For diagnosis it is advantageous to obtain differential diagnostics with which it can be determined whether and to what degree MUC1 is present, which carries tumour-relevant epitopes or less relevant, respectively, irrelevant epitopes, and which does not have a reduced or increased immuno-suppressive effect (see also *infra*, diagnostics). The methods of the invention which are in the following described allow the definition, characterization and isolation of the above MUC1 molecules which are suitable according to the present invention.

The present invention relates to methods allowing the identification and/or generation of such MUC1 molecules or their fragments or derivatives which can advantageously be used in tumour treatment, tumour prophylaxis and tumour diagnosis, from the large pool of different MUC1 molecules, the structure of which is in no case chemically known in detail. Such MUC1 molecules are, according to the invention, designated in the following as tumour-associated MUC1 molecules, which in addition to molecules may also comprise molecule mixtures, such as glycopeptides, glycoproteins or mixtures thereof as well as fragments and derivatives of the molecules containing the tumour epitope of the present invention. According to the present invention, tumour-associated molecules are also cells and other carrier substances, such as viruses, bacteria, parts of cells, such as exosomes or cell lysates, or liposomes, which contain one or more tumour-associated MUC1-molecules with the tumour epitope according to the invention. The methods correspond to screening assays allowing to identify and obtain suitable MUC1 for the applications mentioned above and described below in detail. The central step of the method of production consists in the identification and/or isolation of tumour-associated MUC1 molecules by means of antibodies recognizing the MUC1 tumour epitope according to the present invention, such as antibodies of

group 1 (A76-A/C7, VU-11E2, VU-11D1, BC4E549, VU-12E1, VU-3D1 and b-12). A preferred group of antibodies are such antibodies which show an additive binding effect of length and PDTR glycosylation, such as A76-A/C7, VU-11E2, VU-11D1, BC4E549, VU-12E1, VU-3D1 and b-12. A further preferred group of antibodies are antibodies with a particularly pronounced additive effect, such as A76-A/C7, VU-11E2 and VU-11D1. The antibody A76-A/C7 is preferred according to the present invention for all uses, since it has the most pronounced additive effect. A further antibody which is suitable for the method of production according to the present invention is VU-4H5, which also has a pronounced additive effect, but otherwise is inversely glycosylation-dependent with regard to short PDTR-glycosylated tandem repeats (see Example 4). This group also comprises other antibodies having comparable binding properties.

The individual methods used in the methods are known to the skilled person and, by way of example, are described in detail in the Examples.

In a method according to the present invention, tumour-associated MUC1 molecules are identified and/or isolated and obtained by binding to the above mentioned antibodies.

The methods used for identification are methods known per se, such as ELISA, RIA, Western Blot, Immune Precipitation, Scatchard Blot Analyses, FACS Analyses, MACS Analyses, immunocytochemistry and immunohistology. Most of these methods are described in detail in the Examples.

According to the method of the invention the above described MUC1 molecules may be obtained, as shown above, by affinity chromatography on one or more antibodies from body fluids, such as serum, breast milk or urine, or from supernatants of cell cultures. Further purification and/or concentration steps according to methods known per se can be combined with one or more affinity chromatography steps on the antibodies. Tumour-associated MUC1 molecules can likewise be obtained from tumour cells, tumour tissues or tumour cell lines, by first applying a suitable step according to methods known per se which makes the MUC1 bound to the cell membrane or intracellular MUC1 accessible to affinity purification, for example by solubilization with suitable detergents or by separation of extracellular MUC1 components by proteolysis or cell lysis. A differential purification of the tumour-associated MUC1 molecules from supernatants of cell cultures and from body fluids separated from membrane-bound tumour-associated MUC1 from tumour cells, tumour tissues or tumour cell lines is made possible by a combination of the above described methods. An important step hereby is the separation of supernatants and cells according to methods known per se, such as

centrifugation. For the preparation of secreted and membrane-standing tumour-associated MUC1 molecules, a combination of both methods is used, for example by combining the cell culture supernatants with solubilized, membrane-standing MUC1 before carrying out corresponding affinity chromatography steps on suitable antibodies. Tumour-associated MUC1 molecules according to the present invention also include such that are further purified or first purified by further affinity chromatography steps with other tumour-specific antibodies, for example antibodies or lectins against tumour-associated or tumour-specific carbohydrate antigens, such as the Thomsen-Friedenreich antigen, the Tn antigen, the sialyl-Tn antigen or the Lewis Y-antigen, to generate tumour-associated MUC1 molecules carrying multiple tumour antigens. Such tumour-associated MUC1 molecules are advantageous for tumour therapy, since immune responses against the tumour excited thereby, are more difficult for tumour escape mechanisms to circumvent.

Fragments of tumour-associated MUC1 molecules can be prepared by methods known per se before or after affinity chromatography on suitable antibodies, for example by enzymatic or chemical proteolysis, and purified by suitable methods known per se. Fragmentation may be preceded by partial deglycosylation, for example by one or more enzymatic deglycosylation steps, such as the complete or partial separation of sialic acids by sialidases, or the complete or partial separation of fucose components by one or more chemical deglycosylation steps, such as the per-se known TFMSA method (Trifluoromethylsulfonic acid treatment).

Tumour-associated MUC1 molecules and their fragments may be further modified according to methods known per se. Modifications according to the present invention may for example comprise the modification of the glycosylation in one or more steps, e.g. the enzymatic glycosylation or deglycosylation with glycosyl transferases and/or glycosylidases and/or chemical deglycosylations. The modifications can be made prior to or after affinity chromatography steps on antibodies. The resulting fragments may be further purified or fractionated by suitable purification and/or concentration steps according to methods known per se. Further modifications according to the invention comprise couplings with other substances, e.g. such as increase an immune response against MUC1 on tumour cells and/or favor an application for the treatment and/or prophylaxis of tumours and/or metastases in humans, e.g. coupling of the tumour-associated MUC1 molecules to carrier molecules, such as KLH, or to immunostimulating molecules, such as interleukins, GM-CSF, interferons and/or TNF-alpha, or such as increase the stability of the molecules in-vivo and during storage, or such which increase an expression, or such which cause production-technological advantages, such as

solubility. The couplings are made according to methods known per se, such as chemically via suitable spacers or biochemically in the form of recombinant fusion proteins expressed in cells (see *infra*), such as interleukin 12 or tetanus toxoid peptide.

A method for producing tumour-associated MUC1 molecules according to the present invention is also the synthesis of glycopeptides by methods known per se, which are better bound by the antibodies of the present invention which recognize the MUC1 tumour epitope than the identical glycopeptides which, however, are not glycosylated in the PDTR region. In this connection it may be advantageous to completely or partly synthesize tumour-associated MUC1 molecules obtained by one of the above described methods, and to couple them by one or more additional amino acids, such as cysteine or lysine and carrier molecules, to molecules known per se, such as KLH. A complete or partial analysis is made by methods known per se, for example the fragmenting mass spectrometrical analysis, such as PSD-MALDI-MS or ESI-MS/MS, or NMR or immunochemical methods.

MUC1 glycopeptides according to the present invention are obtained by synthesizing glycopeptides according to methods known per se and testing their binding to MUC1 antibodies. MUC1 glycopeptides bound by antibodies recognizing the MUC1 tumour epitope of the present invention and which bind them in a better way than the identical glycopeptides which, however, are not glycosylated at the PDTR-region correspond to the criteria according to the invention for MUC1 glycopeptides (the mentioned binding properties of these antibodies have already been described *supra* under item ii)). According to the invention such MUC1 glycopeptides are preferred that are better bound by antibodies than corresponding shorter glycopeptides having 1 or 2 tandem repeats but the same glycosylation and the same glycosylation sites (the mentioned binding properties of these antibodies have already been described *supra* under item iv)). In a preferred variant the peptide-chip technology is used. By it a large number of MUC1 peptides is prepared, which are glycosylated on different positions and/or have different lengths. By means of the binding of antibodies to the MUC1 tumour epitope according to the invention, such MUC1 glycopeptides are obtained by the HT method (high throughput) which meet the criteria for MUC1 glycopeptides according to the present invention. In a further preferred variant such MUC1 glycoproteins may be obtained by the same method which in addition are particularly immunogenic for carbohydrate epitopes, by additionally testing the above described peptide-chips with tumour-specific anti-carbohydrate-antibodies and identifying such

glycopeptides which are bound by antibodies to the MUC1 tumour epitope according to the present invention and by antibodies to the tumour-specific carbohydrate antigen. The HT methods are basically known for peptides and may also be used for glycopeptides.

Methods for the preparation of structures carrying several to many tumour-associated MUC1 molecules are carried out in a manner known per se, for example by chemical coupling to liposomes or coupling to lipids and subsequent inclusion in liposomes or by coupling to other carrier structures.

As will be described in the following, also the methods according to the invention also make use of cells or cell lines carrying or secreting, or secreting and carrying MUC1 molecules or fragments thereof which may be prepared by the above described methods. Methods for preparing such cells include the selection of cells which carry the tumour-associated MUC1 molecules by binding to antibodies recognizing the tumour epitope (group 1, preferably A76-A/C7). For this methods are used which are known per se. For example, the cells with tumour-associated MUC1 molecules are obtained by magnetic separation by means of particles binding to a magnet and loaded with the antibodies (MACS sorting) and separated from such cells which carry no or only small amounts of tumour-associated MUC1 molecules. A further example is the sorted extraction of the fluorescence-marked cells carrying the antibodies by means of a FACS sorter. Both methods are known to the skilled person. In this connection it is advantageous to clone by methods known per se the cells obtained by antibody-binding by individualization (limiting dilutions) in suitable media according to methods known per se, in order to obtain stable cell clones and to identify and select via antibody-binding, e.g. by means of quantitative ELISA according to methods known per se, which cell clones secrete, carry or carry and secrete a suitable amount of tumour-associated MUC1 (see Example 5). A part of the production method is preferably a quantitative sandwich ELISA which is explained infra in detail (test method quantitative sandwich ELISA; see Example 5). An alternative method which may also be complementary used is the quantitative determination of the MUC1-positive cells and the amount of tumour-associated MUC1 molecules on the cells via a FACS analysis or a Scatchard blot analysis known per se to the person skilled in the art, by means of the antibodies according to the invention, which recognize the MUC1 tumour antigen.

The cells or cell lines are either used themselves as tumour-associated MUC1 molecules in the applications of the invention or secreted or membrane-standing or secreted and membrane-standing glycoproteins, glycopeptides, or their fragments, are obtained from the cells according to the above described methods of the

invention. Cell lysates, too, are prepared according to the invention by methods known per se, such as by alternative freezing with liquid nitrogen and thawing.

A further method for producing cells or cell lines according to the invention in addition to the above mentioned methods comprises a transformation of cells with recombinant MUC1. For this suitable cells or cell lines are selected and either the expression of MUC1 is increased, novel forms, e.g. fragments, of the MUC1 are expressed or MUC1 is expressed de novo with the aim of obtaining tumour-associated MUC1 molecules or obtaining cells carrying or secreting more or improved tumour-associated MUC1 molecules. For this the MUC1 gene or parts thereof are used. Preferably, the MUC1 gene comprises a DNA sequence selected from the group of sequences deposited at NCBI consisting of: Deposition Nos. NM 002456, XM 053256, AF125525, AF348143, S81781, X80761, X69118, J05582 and M21868. Preferred are such parts of the MUC1 gene which code for shortened versions containing one or more tandem repeats, cloned in a suitable expression vector by methods known per se and introduced into the cells, e.g. by electroporation. The transformed cells are selected according to methods known per se, e.g. via selection with suitable antibiotics, the resistances of which are also encoded on the recombinant genetic material introduced (see Example 5). Stably transformed cells are preferred, as described above, cloned, for example by limiting dilution, preferably in combination with an enrichment of the cells expressing the tumour-associated MUC1 molecules, e.g. by FACS or MACS (see Example 5).

In a further method according to the invention, tumour-associated MUC1-molecule positive cells or cell lines are made sensitive by gene transfer to the treatment of cytostatic drugs, e.g. by the insertion of the HSV-TK gene, which causes the cells to become sensitive for a gancyclovir treatment. The genes are inserted and the transformed cells are selected by methods known per se. According to the present invention, the gancyclovir-sensitive cells are killed by gancyclovir either by their application on humans or after application on humans. This method allows to control living cells or cell lines applied in tumour treatment to humans and is used for this purpose alternatively or complementary to the radiation of cells (see also uses of the present invention).

In a further method according to the present invention, tumour-associated MUC1 molecules are obtained from tissues, by generating the tumour-associated MUC1 molecule by means of the techniques described above from the cell membranes. For this the tissue is broken down by methods known per se to make the membrane-standing tumour-associated MUC1 molecules accessible, e.g. by proteolytic or mechanical methods.

The methods are explained in more detail in the Examples, but are not limited thereto.

By means of the production methods, tumour-associated MUC1 molecules may be obtained which have no or only a reduced immunosuppressive effect and which generate an immune response to the native tumour-MUC1 and to tumour cells. It is shown in Example 5 how tumour-associated MUC1 molecules according to the present invention have no or a reduced immunosuppressive effect. Such tumour-associated MUC1 molecules of the invention are immunogen and can produce an anti-tumour immune response.

Preferably, one part of the production method is a quantitative Sandwich ELISA in which an anti-MUC1 antibody is immobilized, cells or MUC1 obtained from supernatants or membrane preparations are applied in suitable solutions and detected by a second anti-MUC1 antibody. At least one antibody is directed to the tumour-associated MUC1 molecule of the invention. The second antibody is preferably directed to an epitope of MUC1 which does not lie in the PDTR region or to a carbohydrate epitope, such as the Thomsen-Friedenreich antigen. According to the present invention, this quantitative Sandwich ELISA is a test procedure for the preparation of tumour-associated MUC1 molecules, including cells, as well as a test procedure for diagnosis.

To some extent, antibodies to MUC1 are already used for diagnosis and clinically studied for the treatment of tumours. However, the MUC1-specific antibodies recognize different epitopes on MUC1, which has a large number of different epitopes and, moreover, as described above, is not an exactly structurally defined molecule but a heterogeneous mixture of molecules, which differs from tumour to normal tissue, for example in its glycosylation. Thus, at present the HMFG-1 is under clinical study, which does not recognize the MUC1 tumour epitope of the present invention.

The invention shows which antibodies in view of their binding properties are advantageous for the treatment of tumours. Thus the invention allows the selection of better suitable antibodies for the tumour treatment in form of a screening procedure (screening assay): The invention provides a method for the determination of the suitability of antibodies for the treatment of tumours and for diagnosis and for recognizing the tumour epitope of the invention. This method comprises a combination of biochemical tests, which are described in Examples 1 and 2, and preferably also in 3. As described above, antibodies possessing the binding

properties defined in group 1 or of Example 4 in the method, are suitable within the meaning of the invention. By means of this procedure, also antibodies are characterized and obtained which are suitable to determine the MUC1 tumour epitope of the invention and which are suitable to characterize, isolate and obtain tumour-associated MUC1 molecules of the present invention.

Furthermore, the invention provides a method for preparing particularly suitable monoclonal antibodies for therapy and diagnosis of tumour diseases, since they have a particularly pronounced additive effect compared to previously tested antibodies. The method of the invention comprises the immunization of animals or humans with MUC1 material in form of molecules, cells or cell lysates, which material was obtained with the method of the present invention. The antibodies are produced and characterized by methods known per se, including at least one of the above described test systems. The specificity of the antibodies is tested for particularly high additive effects.

Advantageously, the methods of the present invention allow the preparation or identification of tumour-associated MUC1 molecules which are able to produce an immune response in humans, and thus are suitable starting substance for vaccines against tumours which produce tumour-associated MUC-1. The preparation of these MUC1 molecules has only become possible by the methods of the present invention. The present invention is based, inter alia, on the surprising finding that antibodies produced in mice to tumour-associated MUC1 which possess the above described properties are able to specifically bind in the heterogeneous group to such tumour-associated MUC1-molecules which are able to trigger an immune response in humans, as can be shown for example in NOD-SCID mouse test. These MUC1 molecules are preferably characterized by having an increased number of epitopes having an immunostimulating effect and a reduced number of or no immunosuppressive epitopes at all.

The definitions introduced above of the terms are also applicable *mutatis mutandis* for the terms used in the following methods.

The present invention furthermore comprises a method for identifying an MUC1 molecule which is able to trigger an immune response in humans, comprising:

- a) contacting a mixture of MUC1 molecules with an antibody having the following properties:
 - i) binding to the immunodominant region of the MUC1 tandem repeat; and
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- ii) binding to an MUC1 fragment having a length of between 9 and 40 amino acids and containing sequences of the MUC1 tandem repeat and the immunodominant region is made possible or enhanced by glycosylation of the threonine of a PDTR sequence; and
 - iii) binding to unglycosylated MUC1 tandem repeats is enhanced when the unglycosylated MUC1 tandem repeats exist in series; and
 - iv) binding to multiple glycosylated MUC1 tandem repeats carrying glycosylations on the threonines of several PDTR sequences of the immunodominant region is increased due to an additive effect of length and PDTR glycosylation vis-à-vis short MUC1 fragments of ii)
- for a period of time sufficient and under conditions suitable to allow the formation of an immune complex; and
- b) identification of the immune complex.

The term "identifying" within the framework of the invention comprises all measures by which an immune complex can be detected. Suitable procedures are known from the prior art and comprise for example ELISA, MACS, RIA, Scatchard blot analysis, Western blot, immunocytochemistry, immunohistology or FACS methods. Suitable procedures are described in more detail also in the following.

In a preferred embodiment the methods of the invention further comprise at least one of the following steps:

- 1) Obtaining the MUC1 molecule or a mixture thereof in step (a) of the method of the invention from tumour tissue, tumour cells and/or body fluids containing tumour-associated MUC1 molecules, which had been previously isolated;
 - 2) obtaining the MUC1 molecule or a mixture thereof in step (a) of the method of the invention from cells or cell lines expressing and/or secreting tumour-associated MUC1 molecules or mixtures thereof;
 - 3) obtaining the MUC1 molecule or a mixture thereof in step (a) of the method of the invention from recombinant cells or cell lines previously modified by genetic engineering in such a way that they express and/or secrete tumour-associated MUC1 molecules or mixtures thereof;
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- 4) obtaining the MUC1 molecule or a mixture thereof in step (a) of the method of the invention from cell lysates and/or the cell supernatant from tumour tissue, tumour cells and/or body fluids as described under (1) or from cells or cell lines as described under (2) and (3), which contain tumour-associated MUC1.

Suitable tumour tissues or tumour cells can be obtained from tumour patients or persons without tumour, in the latter case, a further treatment, as described elsewhere, may be necessary, such as partial deglycosylation prior to affinity purification.

A preferred embodiment of the present invention are tumour-associated MUC1 molecules in the form of glycoproteins or glycopeptides, which were obtained from the supernatant of cells which secrete a large amount of tumour-associated MUC1 molecules. In the preferred method of the present invention a subcell line of the mamma carcinoma cell line ZR-75-1 by means of ELISA techniques, FACS analyses and immunocytochemical tests identified and characterized as suitable cell line, which express tumour-associated MUC1 molecules on the surface and secrete them into the cell culture supernatant, see also Example 5. Since the expression rate of the individual cells varies, clones (sublines) were prepared from the starting cell line by limiting dilution, which clones carry and/or secrete stably increased amounts of tumour-associated MUC1 molecules, see also Example 5. In a further preferred procedural step of the invention the MUC1 gene or parts thereof were integrated into the cell by an expression vector and procedure more closely described in Example 5 in order to further increase the expression of the secretory tumour-associated MUC1 molecules. The cells were again cloned by limiting dilutions in suitable culture media. From the resulting sublines tumour-associated MUC1 molecules are obtained from the supernatant by means of affinity chromatography on A76-A/C7, which are used in the methods of the invention as tumour agents in tumour treatment and in tumour diagnosis.

In a further preferred procedure, fragments are produced from the obtained tumour-associated molecules by limited proteolysis with clostripain according to methods known per se.

In a further preferred procedure, tumour-associated MUC1 glycopeptides are synthetically produced by methods known per se. These MUC1 glycopeptides are bound to antibodies recognizing the MUC1 tumour epitope of the present invention, and which bind them better than the identical glycopeptides which, however, are not glycosylated on the PDTR region. According to the invention, MUC1 glycopeptides are preferred which are better bound by antibodies than corresponding shorter

glycopeptides having 1 or 2 tandem repeats but the same glycosylation and the same glycosylation sites. The corresponding binding properties have already been described above.

The method of the invention for use on humans comprises suitable pharmaceutical formulations, preferably in combination with adjuvants and/or co-stimulatory molecules, such as GM-CSF and in case of shorter MUC1 glycopeptides a coupling to KLH. A preferred variant is the loading of dendritic cells or cells possessing an immunostimulatory function comparable to dendritic cells, with cell lysates, or the fusion of cells carrying the tumour-associated MUC1 molecule I with dendritic cells according to procedures known per se, and the administration of these cells, preferably after radiation, to humans according to methods known per se. The treatment with loaded dendritic cells and glycopeptides and/or glycoproteins is preferably combined, later boosting being preferably carried out with the glycopeptides and/or glycoproteins of the invention in suitable pharmacological formulations according to the invention, preferably with adjuvants or co-stimulatory molecules (such as GM-CSF).

The preferred use according to the invention of the glycopeptides and glycoproteins or of the dendritic cells loaded with glycopeptides and/or glycoproteins is for the treatment and prophylaxis of MUC1-positive tumour diseases, such as mamma carcinoma, ovarian carcinoma, colon carcinoma, gastrointestinal carcinoma, pancreas carcinoma, lung carcinoma, multiple myeloma, and against primary tumours, minimal residual tumour diseases, metastases and as adjuvant treatment. Preferably, the glycopeptides and glycoproteins or the dendritic cells loaded with glycopeptides and/or glycoproteins are administered subcutaneously, intradermally, intrarectally, intranodally (lymph nodes) or systemically by procedures known per se.

The present invention also relates to a method for producing cells which comprise an MUC1 molecule capable of generating an immune response in humans, comprising:

- a) contacting a mixture of cells containing MUC1 molecules with an antibody having the following properties:
 - i) binding to the immunodominant region of the MUC1 tandem repeat, and
 - ii) binding to an MUC1 fragment having a length of between 9 and 40 amino acids and containing sequences of the MUC1 tandem repeat and the
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- immunodominant region is made possible or enhanced by glycosylation of the threonine of a PDTR sequence; and
- iii) binding to unglycosylated MUC1 tandem repeats is enhanced when the unglycosylated MUC1 tandem repeats exist in series; and
 - iv) binding to multiple glycosylated MUC1 tandem repeats carrying glycosylations on the threonines of several PDTR sequences of the immunodominant region is increased due to an additive effect of length and PDTR glycosylation compared to short MUC1 fragments of ii)
- for a period of time sufficient and under conditions suitable to allow the formation of an immune complex;
- b) isolation of the cells which have formed an immune complex; and
 - c) making the cells from the immune complex available.

The term "cells which comprise an MUC1 molecule capable of generating an immune response in humans" comprises all cells which produce a tumour-associated MUC1 molecule as described above. Such cells are preferably cells which can be isolated or prepared from tumour tissue, tumour cell lines, or cells or cell lines, modified by genetic engineering in such a way that they produce a tumour-associated MUC1 molecule as described above. The last mentioned cells or cell lines may be produced by methods known from the prior art. These methods comprise for example procedures with which foreign DNA can be stably or transiently inserted into the cells or cell lines. In the following, further suitable procedures are also described in more detail.

The term "isolation of the cells" means all measures for the separation of cells which have formed an immune complex with antibodies due to the tumour-associated MUC1 molecules produced by them. The skilled person knows these methods. The preferred method for this is the FACS or the MACS method. In the following, further suitable methods are described in more detail.

The previously introduced definitions of the terms are applicable *mutatis mutandis* to the terms used in the below described methods.

The present invention further comprises a method for the identification of cells comprising an MUC1 molecule capable of generating an immune response in humans, comprising:

- a) contacting a mixture of cells comprising MUC1 molecules with an antibody having the following properties:
 - i) binding to the immunodominant region of the MUC1 tandem repeat, and
 - ii) binding to an MUC1 fragment having a length of between 9 and 40 amino acids and containing sequences of the MUC1 tandem repeat and the immunodominant region, is made possible or enhanced by glycosylation of the threonine of a PDTR sequence; and
 - iii) binding to unglycosylated MUC1 tandem repeats is enhanced when the unglycosylated MUC1 tandem repeats exist in series; and
 - iv) binding to multiple glycosylated MUC1 tandem repeats carrying glycosylations on the threonines of several PDTR sequences of the immunodominant region, is increased due to an additive effect of length and PDTR glycosylation as compared to short MUC1 fragments of ii) for a period of time sufficient and under conditions suitable to allow the formation of an immune complex; and
- b) identification of the immune complex.

In a preferred embodiment the methods of the invention further comprise at least one of the following steps:

- 1) Obtaining of cells, cell lines or subcell lines carrying and/or secreting tumour-associated MUC1 molecules or mixtures thereof in step (a) of the method of the invention from tumour tissue or tumour cells containing tumour-associated MUC1 molecules, which had been previously isolated, with or without subsequent cell cloning;
 - 2) obtaining of cells, cell lines or subcell lines carrying and/or secreting tumour-associated MUC1 molecules or mixtures thereof in step (a) of the method of the invention from cells or cell lines comprising tumour-associated MUC1 molecules, with or without subsequent cell cloning;
 - 3) obtaining of cells, cell lines or subcell lines carrying and/or secreting tumour-associated MUC1 molecules or mixtures thereof in step (a) of the method of the invention from recombinant cells or cell lines previously modified by genetic engineering in such a way that they express and/or secrete tumour-associated MUC1 molecules or mixtures thereof, with or without subsequent cell cloning;
 - 4) obtaining of cells, cell lines or subcell lines carrying and/or secreting tumour-associated MUC1 molecules or mixtures thereof in step (a) of the method of the invention, which were modified by genetic engineering in such a way that
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they carry and/or secrete immunostimulatory molecules, with or without subsequent cell cloning;

- 5) obtaining of cell lysates or mixtures of cell lysates from cells, cell lines or subcell lines as described under (1) to (4) above, which contain tumour-associated MUC1.

In a further preferred embodiment of the methods for the present invention, the antibody is selected from the group comprising A76-A/C7, VU-11E2, VU-11D1, BC4E549, VU-12E1, VU-3D1 and b-12.

Particularly preferred is the antibody A76-A/C7 and recombinant forms thereof, as will be described below in more detail. In a further embodiment of the method of the present invention the antibody is VU-4H5.

The antibody A76-A/C7 recognizes the MUC1 tumour epitope and has a particularly strong additive effect. Compared to the HMFG-1 which is undergoing clinical development, the A76-A/C7 according to the invention has a number of advantages: The A76-A/C7 recognizes the MUC1 tumour epitope of the present invention and was prepared as immunogen by means of a desialinated mamma carcinoma cell line (T47D). In contrast, HMFG-1 does not recognize the MUC1 tumour epitope of the invention and was prepared as immunogen by means of MUC1 from human milk (see also Examples 1 to 3). A76-A/C7 for example clearly distinguishes normal tissue and benign adenoma (negative) from malignant colon tumours such as carcinoma in-situ, carcinoma and metastases (strongly positive), whereas HMFG-1 does not make this clear differentiation. A76-A/C7 is a good serum marker for tumour diagnosis, whereas HMFG-1 is not suitable as serum tumour marker to the same degree. Moreover, the affinity of the A76-A/C7 is higher by a multiple than that of HMFG-1. Thus, compared to the HMFG-1 presently in clinical development, A76-A/C7 constitutes a significant improvement for applications according to the present invention.

The present invention also relates to a method for preparing an antibody comprising:

- (a) carrying out the steps of the production and identification methods described above;
 - (b) inserting the MUC1 molecule, the cell or the cell lysates into an animal; and
 - (c) providing an antibody specifically recognizing the MUC1 molecule and showing a particularly pronounced additive effect.
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In this context the expression “particularly pronounced additive effect” is to be understood as meaning that the increase of the binding of the antibody to multiple PDTR-glycosylated tandem repeats as described under 4) compared to short PDTR-glycosylated MUC1 peptides as described under 2), and both compared to not PDTR-glycosylated short MUC1 peptides is stronger than the increases of the binding of the antibodies A76-A/C7, VU-11E2, VU-11D1, BC4E549, VU-12E1, VU-3D1 and b-12 (or comparable to the strongest increases of these antibodies).

The definitions of the terms given above apply *mutatis mutandis* to these and the following embodiments.

The term “antibodies” as used according to the present invention comprises:

- murine, chimeric, humanised, human antibodies
- whole antibodies of various isotypes or antibody fragments, such as single-chain antibody fragments (ScFv), Fab fragments, multivalent antibody fragments (e.g. diabodies, triabodies, tetrabodies), or antibody fragments of other formats, including those used as fusion proteins coupled with other peptides or protein structures.
- antibodies or antibody fragments which beside the specificity for MUC1 according to the present invention also possess one or more further specificities, e.g. bispecific antibodies.
- molecules which strictly speaking are no antibodies, but which possess binding properties identical to those of the antibodies according to the present invention which carry the MUC1 tumour epitope, and/or parts of antibodies, such as CDR regions or fragments thereof, such as affibodies or other carrier structures carrying binding domains for specific binding. The invention also relates to antibodies derived from the antibodies here described and which thus acquire, increase an additive binding effect and/or bind better to the multiple PDTR-glycosylated tandem repeats than the starting antibody. The corresponding derivation of the antibody may result from methods known per se by deliberate or accidental modifications of the antibody and is tested by means of the comparing binding to corresponding MUC1 glycopeptides as described under Examples 1 to 3.

These antibodies are provided by techniques known per se. The antibodies may be obtained as polyclonal antibodies directly from the serum of the animals or by the known methods for producing monoclonal antibodies as such (Example 6).

The antibodies prepared by the method of the invention may advantageously be used against the MUC1 tumour epitope for the treatment of tumour diseases. These antibodies can be directly utilized in the treatment or prophylaxis of tumour diseases according to the present invention or coupled with effector structures. Effector structures according to the invention mean such chemical or biochemical compounds, molecules or atoms, which directly or indirectly cause the killing or harming, including for example the slowdown or inhibition of the growth of tumour cells. These include for example: radioisotopes, toxins, cytostatica and other effector molecules, such as cytokines and chemokines or other structures which themselves are effectors or are coupled to the effector molecules, for example liposomes loaded with toxins or cytostatica, which carry MUC1 antibodies. In the latter example, also such effector structures are meant which in addition to the MUC1 antibody for the tumour specificity also carry such molecules which are responsible for the reception of the effector structures or fragments thereof in the cells, such as antibodies to receptors causing receptor-mediated endocytosis and thus improving the MUC1-mediated reception. The combination of the antibodies with the effector structures is effected by methods known per se. The couplings may for example be directly effected by covalent or non-covalent loading, by chemical coupling, in which an additional chemical or biological molecule may be necessary, such as a chelator or a linker, or by fusion in the form of fusion proteins or fusion peptides.

The antibodies are used in the treatment of tumour diseases with MUC1-carrying tumour cells or for prophylaxis, which for example prevents the formation of primary tumours or metastases. Here, the preferred goal is the treatment of the minimal residual diseases and of metastases. The antibodies of the invention are administered in a suitable formulation one time or repeatedly at suitable intervals and dosages. A preferred form are radioactively labelled antibodies. A further preferred form are radioactively labelled diabodies, triabodies and tetrabodies. Further preferred forms are radioactively labelled single chain-antibody fragments and chimeric antibodies. However, the invention is not limited to these antibodies, the radioactive labelling and these antibody formats.

In the method of the invention for tumour diagnosis and prognosis, MUC1-specific antibodies recognizing the tumour epitope of the invention are utilized in methods known per se to detect MUC1 in serum or in tissue preparations. According to the invention free MUC1, MUC1 present in immune complexes and MUC1 bound on cells is detected and the presence of the tumour-associated MUC1 molecules is determined qualitatively, quantitatively and/or in relative quantities by methods

known per se. The same methods are used according to the invention also for the development control of tumour diseases and for the control of the development of the treatment. The methods used in the procedures are known per se, e.g. ELISA, Western blot, FACS (fluorescence-activated cell sorting), MACS (magnetic cell sorting), ADCC (antibody-dependent cell-mediated cytotoxicity), CDC (complement-dependent cytotoxicity), immunocytochemistry and immunohistochemistry. An example for this is the Sandwich ELISA of the invention described further below with the antibodies described there and methods described in the examples. In a preferred method, antibodies, which recognize the tumour epitope of the invention, are compared with such antibodies which do not recognize the tumour epitope, to improve differentiation and evaluation. This serves to diagnose MUC1-positive tumours, to monitor the course of the tumour disease and to make a prognosis. The binding of the antibodies also determines the ratio of immunosuppressive MUC1, since MUC1 which is bound by antibodies against the tumour epitope of the invention does not have any or only a reduced immunosuppressive effect and in suitable formulations can specifically activate the immune system against tumour-MUC1.

In a method of the invention the antibodies recognizing the tumour epitope of the invention and such as are prepared by the method of the invention are used for in-vivo diagnosis. For this the antibodies are labelled by suitable methods known per se and are thus made available for imaging methods on humans known per se, such as radioimmunodiagnosis, PET scan process or immunofluorescendoscopy, for example by coupling and/or loading with corresponding molecules, such as radioactive isotopes, e.g. indium, or fluorescent dye stuffs such as Cy3, Cy2, Cy5 or FITC.

The present invention further relates to a method for producing a pharmaceutical composition comprising the steps of the method of the invention and further comprising the steps of formulating the MUC1 molecule, the cell, the cell lysates or the antibodies or combinations thereof in pharmaceutically acceptable form.

The term "pharmaceutical composition" according to the invention defines substances and preparations of substances intended to heal, relieve or prevent by application on or in the human body, diseases, suffering, physical injuries or abnormal complaints. During the production process of the invention, medicinal and/or pharmaco-technical excipients may be added to the compounds identified by the methods of the invention. According to the present invention, medicinal

excipients are such substances which are used for the production (as active ingredients) of pharmaceutical compositions in a method according to the present invention. Pharmaco-technical excipients merely serve to suitably formulate the pharmaceutical and may even be subsequently removed if they are only necessary during the process, or they may form a part of the pharmaceutical composition as pharmaceutically acceptable carriers. Examples for pharmaceutically acceptable carriers are given below.

The pharmaceutical may be formulated optionally in combination with a pharmaceutically acceptable carrier and/or diluent.

Examples for suitable pharmaceutically acceptable carriers are known to the skilled person and comprise phosphate-buffered saline solutions, water, emulsions, such as oil/water emulsions, different kinds of detergents, sterile solutions, etc. Pharmaceutical compositions comprising such carriers can be formulated by conventional methods. These pharmaceutical compositions can be administered to an individual in a suitable dosage, e.g. in a range from 1 μ g to 100 mg per day and patient. The administration may be carried out in different ways, e.g. directly to the skin, intravenously, intraperitoneally, subcutaneously, intramuscularly, locally or intradermally. The administration of nucleic acids can also be made in the form of gene therapy. The kind of dosage is decided by the physician in charge depending on the clinical factors. The skilled person knows that the type of dosage depends on various factors such as the size, the body surface, the age, the gender or the general health of the patient, but also depending on the specific drug which is administered, the duration and kind of administration and on other medicaments which may possibly be administered in parallel.

Tumour pharmaceuticals according to the present invention comprise a pharmacological substance containing one or more tumour-associated MUC1 molecules according to the invention, cells comprising these or cell lysates prepared from these cells in a suitable solution or dosage form. These may be administered either alone or in combination with one or more adjuvants or another suitable substance for intensifying the effectiveness. The preferred adjuvants used are QS-21, GPI-0100 or other saponines, water-oil emulsions, such as montanide adjuvants, polylysin, polyarginine compounds, DNA compounds, such as CpG, Detox, bacterial vaccines, such as typhus vaccine or BCG vaccines, and are mixed in a suitable manner according to procedures known per se with the tumour-associated MUC1 molecules of the invention and/or with the antibodies. The tumour pharmaceuticals are produced by methods known per se. According to the present invention a tumour pharmaceutical is also a combination of 2 or more of the tumour

pharmaceuticals of the invention, as well as a combination with other tumour vaccines or tumour treatments, such as antibody therapies, chemotherapies or radiotherapies, which in a suitable manner may be administered or used simultaneously or separately. The tumour pharmaceuticals are prepared by methods known per se.

According to the present invention, the tumour pharmaceuticals are used for the treatment of MUC1-positive tumour diseases, such as mamma carcinomas, ovarian carcinomas, colon carcinomas, gastrointestinal carcinomas, pancreas carcinomas, lung carcinomas, multiple myeloma. The treatment may also be an adjuvant treatment for example against primary tumours, minimal residual tumour diseases, metastases. According to the present invention, the tumour pharmaceuticals are used also for the prophylaxis of MUC1-positive tumour diseases. The prophylactic use aims for example to the prophylaxis of the tumour as well as of metastases. The tumour pharmaceuticals are administered in a suitable form by methods known per se. A preferred variant is the injection or administration of the tumour vaccines subcutaneously, intradermally, systemically, intravenously, intraperitoneally, intrarectally, locally into body cavities, such as the peritoneum, or locally directly into organs or lymph nodes. Preferably, different forms of administration may also be combined, wherein they can be administered on different days of treatment or on one treatment day. According to the invention, also 2 or more of the tumour pharmaceuticals of the invention may be combined or one or more of the tumour pharmaceuticals of the invention with one or more tumour pharmaceuticals or tumour treatments, such as antibody therapies, chemotherapies or radiotherapies which can be administered or used simultaneously or separately.

The method of the invention also comprises the production of a tumour pharmaceutical containing a cell vaccine obtainable by the loading of dendritic cells according to methods known per se with the tumour-associated MUC1 molecules of the present invention, and optionally by subsequent maturing of the cells. The resulting cell vaccine may be administered according to methods known per se. For example, to immature dendritic cells are added in a manner known per se tumour-associated MUC1 molecules. The dendritic cells receive the tumour-associated MUC1 molecules, process them and present fragments thereof on their surface in context with MHC molecules and co-stimulatory molecules. After a further maturing according to methods known per se, the cells are used on humans in a suitable formulation. A further example is the loading of the mature dendritic cells according to the known methods of pulsing. Dendritic cells are autologous, allogenic or

semiallogenic dendritic cells or their precursor cells or cells from cell lines possessing the functional properties of dendritic cells, which are subjected *ex vivo* according to methods known *per se* to a suitable treatment for their development and maturing. The preparation of these tumour vaccines with suitable dendritic cells and MUC1 molecules according to the invention, cells or cell lysates, is carried out by methods known *per se* (see Example 7). According to the present invention, a further variant of the tumour pharmaceuticals consists in a T-cell therapy, in which T-cells recognizing suitable tumour-associated MUC1 molecules according to the invention, including fragments or processed derivatives thereof, which correspond to those by processing and presentation *in vivo*, in the context with MHC class molecules and co-stimulatory molecules and which are activated during presentation via suitable antigen-presenting cells. The preparation of such T-cell therapies is carried out according to methods known *per se* (see Example 7).

In a particularly preferred embodiment of the method of the present invention, cell lysates of cells enriched for tumour-associated MUC1 molecules on the cells, are prepared and utilized in the here described applications in tumour treatment and tumour prophylaxis. The methods of the invention for use on humans comprise suitable pharmaceutical formulations, preferably in combination with adjuvants and/or co-stimulatory molecules, such as GM-CSF. Here, a preferred variant is the loading of dendritic cells or cells possessing an immunostimulatory function like dendritic cells, with cell lysates, and the administration of these cells, preferably after radiation, to humans, according to methods known *per se*. The treatment with loaded dendritic cells and cell lysates is preferably combined, with later boosterings preferably carried out with cell lysates in suitable pharmaceutical formulations of the invention, preferably with adjuvants or co-stimulatory molecules (such as GM-CSF).

According to the invention, the preferred use of the cell lysates or the dendritic cells loaded with cell lysates is for the treatment and prophylaxis of MUC1-positive tumour diseases, such as mamma carcinomas, ovarian carcinomas, colon carcinomas, gastrointestinal carcinomas, pancreas carcinomas, lung carcinoma, multiple myeloma, and thereby against primary tumours, minimal residual tumour diseases, metastases, and as adjuvant treatment. The cell lysates or the dendritic cells loaded with cell lysates are preferably administered subcutaneously, intradermally, intrarectally, intranodally (lymph nodes) or systemically by methods known *per se*.

The definitions of the terms introduced above may be applied *mutatis mutandis* to the terms used in connection with the following embodiments.

The present invention furthermore relates to a method of preparing a diagnostic drug comprising the steps of the methods of the invention and further comprising the step of the formulation of the MUC1 molecule, of the cell, of the cell lysates or the antibodies in a diagnostically usable form.

According to the invention, the term "diagnostic drug" defines substances and preparations from substances aimed at recognizing by application on or in the human body or parts thereof, diseases, sufferings, physical injuries or abnormal complaints. The parts of the human body preferably mean body tissue samples or body fluids, such as blood, lymph, urine, spinal fluid or semen, or tissue biopsies.

The formulation of the diagnostic drug preferably comprises the modification of the prepared MUC1 molecules with substances allowing the detection of the molecule. Suitable substances are known from the prior art. On the basis of the chosen substance, the person skilled in the art is able to use suitable measures for formulating the diagnostic drug.

According to the invention, for the diagnosis, substances can also be coupled to the tumour-associated MUC1 molecules according to methods known as such, which facilitate the detection of MUC1-specific antibodies, for example by means of biotinylation of the tumour-associated MUC1 molecules and subsequent immobilisation on ELISA plates.

In a further method for the diagnosis and prognosis of tumours according to the invention, tumour-associated MUC1 antibodies according to the invention are used which carry the tumour epitope, for the determination of MUC1 antibodies in the serum of humans, which recognise the tumour epitope. In a preferred method, MUC1 molecules or fragments thereof carrying the tumour epitope of the invention are used in comparison with antibodies not carrying the tumour epitope in order to achieve a better differentiation and assessment. In this context, according to the invention, free antibodies and antibodies present in immune complexes are detected and their specificities against the MUC1 tumour epitope according to the invention and, preferably other epitopes on the MUC1, including carbohydrate antigens, are determined qualitatively, quantitatively and/or in relative quantities according to methods known *per se*. According to the invention, the same methods are also used to monitor the course of tumour diseases and to monitor the course of

treatments including monitoring of immune responses, for example IgG- and IgM-antibody responses against different epitopes including the MUC1 tumour epitope on MUC1, for the control and dosage of tumour treatment. The methods used in the processes are known per se, for example ELISA, Western blot, FACS (fluorescence-activated cell sorting), MACS (magnet cell sorting), ADCC (antibody-dependent cell-mediated cytotoxicity), CDC (complement-dependent cytotoxicity), immunocytochemistry and immunohistochemistry. One example thereof are glycopeptides coupled to a carrier molecule or biotin immobilised in an ELISA, where the bond of serum antibodies from humans is specifically detected, quantified and thus, assessed in their specificity by means of anti-human-IgG or anti-human-IgM according to a detection method known as such. Such a method is used, for example, to determine the natural antibody titre of MUC1-specific antibodies in the serum and, at the same time, to subdivide it into the different specificities. This is suitable, for example, for the tumour prognosis of breast cancer. Female patients with a high titre of antibodies against the MUC1 tumour epitope according to the invention have clearly improved prospects compared to patients with a lower titre against the MUC1 tumour epitope according to the invention.

The definitions of the terms introduced above can be applied *mutatis mutandis* for the terms in the embodiments described below.

Furthermore, the present invention relates to the use of a MUC1 molecule, a cell, a cell lysate or an antibody obtainable by means of a method according to the invention for the production of a pharmaceutical composition for the prevention or treatment of tumours.

Preferably, within the meaning of the use according to the invention, the pharmaceutical composition is a vaccine.

From the explanation given above it follows that the present invention further relates to the use of an antibody obtainable by means of a method according to the invention for the production of a diagnostic means for the diagnosis of tumours.

Finally, the present invention comprises a purified tumour-associated MUC1 molecule having an immunostimulatory effect in humans and, thus, obtainable by means of a method according to the invention.

The term "purified" defines a MUC1 molecule which can be produced by means of the method according to the invention. Thus, a purified tumour-associated MUC1

molecule also comprises a group of specific MUC1 glycoproteins which are characterised by the fact that they are able to trigger an immune response in humans, however, have only little or preferably no immunosuppressive effect. Further functional and structural characteristics of the purified MUC1 molecule according to the invention are also described in the examples below.

The figures show:

Figure 1 shows examples for the binding behaviour of different anti-MUC1 antibodies. In total, 4 behaviour patterns were defined:

- 1) GD-1: Antibodies binding to the PDTR motif only after the glycosylation thereof (GD refers to glycosylation-dependent). Example: A76-A/C7 (Fig. 1A).
- 2) GD-2: Antibodies whose linkage is improved by glycosylation at the PDTR motif. Example: VU-3D1 (Fig. 1B).
- 3) iGD: Antibodies whose linkage is reduced or prevented (inverse glycosylation-dependency). Example: VU-4H5 (Fig. 1B).
- 4) GI: Antibodies whose linkage to the PDTR region is independent from the glycosylation thereof (glycosylation-independent). Example: HmFG-1 (Fig. 1B).

Figure 2 shows examples for the binding behaviour of anti-MUC1 antibodies towards non-glycosylated MUC1 peptides of different lengths (1-6 tandem repeats).

The results show that antibodies which were produced by an immunisation of mice with tumour material

- positively depend on the glycosylation in the PDTR region of short MUC1 peptides (GD-1 and GD-2)
- and
- positively depend on the increasing length of non-glycosylated MUC1 peptides and, thus, on the number of tandem repeats;

antibodies which were produced by an immunisation of mice with non-tumour

material are

- independent from (GI) or negatively dependent on (inversely dependent; iGD) the glycosylation in the PDTR region of short MUC1 peptides (GI)
or
- independent from the length of the MUC1 peptides and, thus, the number of tandem repeats.

Figure 3 shows 2 examples for the binding of MUC1-specific monoclonal antibodies to glycosylated MUC1 peptides of different lengths (= different number of tandem repeats).

Figure 4 shows the binding of the antibody VU-4H5 to glycosylated MUC1 peptides of different lengths.

Figure 5A: Immunocytochemical staining of ZR-75-1 cells (1) and cells of the subcell line (2) with the antibody A76-A/C7.

Figure 5B: Secretion of A76-A/C7 positive MUC1 from ZR-75-1 sub and MCF-7 cells. The secreted MUC1 was detected 3, 4, 5, 6 and 7 days after spreading 10^5 cells in the cell culture supernatant by means of ELISA with the HMFG1 and A76-A/C7 antibodies. Attention has to be paid to the different dilutions of the cell culture supernatants.

Figure 5C: Expression vector for the integration of the MUC1 cDNA under the control of the CMV promoter in the ZR-75-1 genome as well as the MUC1 cDNA sequence.

Figure 5D: Rec site vector for the integration of the recombination site in the ZR-75-1 genome and illustration of the method of the integration.

Figure 5E: SDS-polyacrylamide gel analysis of different MUC1 preparations.

Figure 5F: The purified secreted tumour-associated MUC1 show no immunosuppressive effects on T-cells *in vitro*.

Figure 6: Stimulation of naïve T-cells by means of the effective dendritic cells charged with the MUC1 of the invention and their MUC1-specific anti-tumour activity (details see exemplary text).

The invention is now illustrated by the following examples. The examples only serve the purpose of explanation and do not limit the scope of protection of the invention.

Example 1: Binding behaviour of different anti-MUC1 antibodies to a MUC1 peptide dependent on the glycosylation of the PDTR motif

Monoclonal anti-MUC1 antibodies (mouse) were tested in the enzyme immunoassay (ELISA) as to their binding capability towards synthetic MUC1 peptides and glycopeptides with the following sequence: biotin-APPAHGVTSAPDT(GalNAc)RPAPGSTAPPAHGVTS. The same but non-glycosylated peptide served as control. The antigens were immobilised on streptavidin-coated microtestplates (BioTeZ, Berlin) at a concentration of 0.5 µg/ml (100µl/well). After washing them three times with PBS/0.05 % Tween 20, various purified anti-MUC1 antibodies were applied in dilution series and incubated at 37°C for 2 hours. Rabbit-anti-mouse-Ig, peroxidase-marked (Dako, Hamburg), served as the second antibody in a dilution of 1:4,000 (1.5 hrs, 37°C). After washing it three times, the plate was developed with ortho-phenylenediamine and the colour reaction was stopped with 2.5 N sulfuric acid. The optical density was measured at 492 nm.

Figure 1 shows examples for the binding behaviour of different anti-MUC1 antibodies. In total, 4 behaviour patterns were observed:

- 1) GD-1: Antibodies binding to the PDTR motif only after the glycosylation thereof (GD refers to glycosylation-dependent). Example: A76-A/C7 (Fig. 1A).
 - 2) GD-2: Antibodies whose linkage is improved by glycosylation at the PDTR motif. Example: VU-3D1 (Fig. 1B).
 - 3) iGD: Antibodies whose linkage is reduced or prevented by glycosylation at
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the PDTR motif (inverse glycosylation-dependency). Example: VU-4H5 (Fig. 1B).

- 4) GI: Antibodies whose linkage to the PDTR region is independent from the glycosylation thereof (glycosylation-independent). Example: HMFG-1 (Fig. 1B).

Table 1 contains an overview of the binding patterns of a panel of monoclonal anti-PDTR (MUC1) antibodies with PDTR-glycosylated and non-glycosylated MUC1 peptides. The table is divided in two sub-tables (A) and (B). Table 1A contains antibodies which were produced by an immunisation with MUC1 from tumour material. Table 1B contains antibodies which were produced by an immunisation with MUC1 from non-tumour material.

Table 1:

Antibodies against the immunodominant PDTR region of the MUC1:

Binding behaviour towards MUC1 peptides glycosylated at the PDTR and non-glycosylated dependent on the type of the immunogen

A. MUC1 from tumour material

| Antibody | immunogen | binding pattern | | | |
|----------|--|-----------------|------|-----|----|
| | | GD-1 | GD-2 | iGD | GI |
| A76-A/C7 | tumour cell line T-47D | + | | | |
| VU-11E2 | tumour cell line ZR75-1 | + | | | |
| VU-11D1 | tumour cell line ZR75-1 | + | | | |
| Ma552 | tumour cell line ZR75-1 | | + | | |
| VU-3C6 | tumour cell line ZR75-1 | | + | | |
| BC4E549 | membrane preparation of the tumour cell line T-47D | | + | | |
| VU-12E1 | tumour cell line ZR75-1 | | + | | |
| VU-3D1 | tumour cell line ZR75-1 | | + | | |
| b-12 | mixture of several tumour cell lines | | + | | |
| B27.29 | MUC1 from tumour Ascites | | + | | |
| MF 06 | MUC1 from ovarian cyst | | + | | |

B. MUC1 peptides (non-glycosylated) or preparations from non-tumour material

| Antibody | immunogen | binding pattern | | | |
|----------|----------------------------------|-----------------|------|-----|----|
| | | GD-1 | GD-2 | iGD | GI |
| VU-4H5 | 60-mer, BSA conjugated | | | + | |
| HMPV | HMFG from breast milk | | | + | |
| HMFG-1 | HMFG from breast milk | | | | + |
| BC2 | HMFG from breast milk | | | | + |
| VA2 | 100-mer fusion protein | | | | + |
| Mc5 | HMFG from breast milk | | | | + |
| E29 | HMFG from breast milk | | | | + |
| BC3 | HMFG from breast milk | | | | + |
| 214D4 | MUC1 peptide (fusion protein) | | | | + |
| BCP8 | MUC1 peptide | | | | + |
| VA1 | 100-mer fusion protein | | | | + |
| C595 | MUC1 from urine | | | | + |
| BC4W154 | HMFG from breast milk | | + | | |

Example 2: Binding of MUC1-specific antibodies towards the immunodominant PDTR region to non-glycosylated MUC1 peptides of different lengths (non-glycosylated multiple tandem repeats)

The MUC1 peptides of the following sequence of different lengths (oligomerisation)

$[VTSPDTRPAPGSTAPPAHG]_n$; $n = 1-6$

were immobilised by drying on a 96-well-TC-microtitre plate at a concentration of 0.5 µg/well in coating buffer at 37°C overnight. The plates were washed (PBS/0.05% Tween20) and subsequently incubated with 50 µl/well purified antibodies at a concentration of 10 µg/ml for 2 hrs. After washing the plate three times, it was incubated with peroxidase-marked polyclonal rabbit-anti-mouse immunoglobulin serum (P260, Dako) in a dilution of 1:2,000. After washing the plate three times, it was developed with ortho-phenylenediamine and the colour reaction was stopped with 2.5 N sulfuric acid. The optical density was measured at 492 nm.

The number of tandem repeats per solution was kept constant in the experiments by using, per well, the same amounts in weight for all the peptides compared.

The results are summarised in Table 2:

The classification was carried out according to the ratio of the optical density 100-mer/20-mer = X as follows:

Without significant bond to the 20-mer, however, a clearer bond to the 100-mer: group LD1 (absolutely dependent on length)

$X > 3.0$: group LD2 (strongly dependent on length)

$X = 1.5 - 3.0$: group LD3 (moderately dependent on length)

$X < 1.5$: group LI (independent from length).

Furthermore, the data about the glycosylation dependency from Table 1 have been included in the table for comparison.

Table 2:

Binding behaviour of monoclonal anti-MUC1 antibodies against the immunodominant PDTR motif: Binding behaviour towards MUC1 peptides glycosylated at the PDTR and non-glycosylated, dependent on the type of the immunogen and with non-glycosylated MUC1 peptides of different lengths (1-6 tandem repeats).

Binding behaviour of the antibodies dependent on:

A. MUC1 from tumour material

| | glycosylation | | | | length | | | |
|----------|---------------|------|-----|----|--------|------|------|----|
| | GD-1 | GD-2 | iGD | GI | LD-1 | LD-2 | LD-3 | LI |
| A76-A/C7 | + | | | | + | | | |
| VU-11E2 | + | | | | + | | | |
| VU-11D1 | + | | | | + | | | |
| Ma552 | | + | | | + | | | |
| VU-3C6 | | + | | | + | | | |
| BC4E549 | | + | | | | + | | |
| VU-12E1 | | + | | | | + | | |
| VU-3D1 | | + | | | | | + | |
| b-12 | | + | | | | | + | |
| B27.29 | | + | | | | | + | |
| MF 06 | | + | | | | | + | |

B. MUC1 peptides (non-glycosylated) or preparations from non-tumour material

| | glycosylation | | | | length | | | |
|---------|---------------|------|-----|----|--------|------|------|----|
| | GD-1 | GD-2 | iGD | GI | LD-1 | LD-2 | LD-3 | LI |
| VU-4H5 | | | + | | | + | | |
| HMPV | | | + | | | + | | |
| HMFG-1 | | | | + | | + | | |
| BC2 | | | | + | | + | | |
| VA2 | | | | + | | + | | |
| Mc5 | | | | + | | + | | |
| E29 | | | | + | | | + | |
| BC3 | | | | + | | | + | |
| 214D4 | | | | + | | | + | |
| BCP8 | | | | + | | | + | |
| VA1 | | | | + | | | + | |
| C595 | | | | + | | | | + |
| BC4W154 | | + | | | | | | + |

Figure 2 shows examples for binding behaviour of anti-MUC1 antibodies towards non-glycosylated MUC1 peptides of different lengths (1-6 tandem repeats).

Example 3: Binding of MUC1-specific antibodies to multiple tandem repeats glycosylated to PDTR

Synthetic MUC1 glycopeptides of the type:

[AHGVTSAPDT(GalNAc)RPAPGSTAPPA]_n; n = 1-5

were immobilised by drying on a 96-well-TC-microtestplate at a concentration of 0.5 µg/well in water at 4°C overnight. The plates were washed (PBS with 0.05% Tween20) and subsequently incubated with 50 µl/well of a solution of 10µg/ml purified antibody in PBS/BSA at 37°C for 2 hrs. After washing the plate three times, it was incubated with peroxidase-marked polyclonal rabbit-anti-mouse immunoglobulin serum (Dako) in a dilution of 1:2,000 at 37°C for 1,5 hrs. After washing the plate three times, it was developed with ortho-phenylenediamine and the colour reaction was stopped with 2.5 N sulfuric acid. The optical density was measured at 492 nm.

The number of tandem repeats per solution was kept constant in the experiments by using the same amounts in weight for all the glycopeptides compared per well.

Figure 3 shows 2 examples for the linkage of MUC1-specific monoclonal antibodies to glycosylated MUC1 peptides of different lengths (= different number of tandem repeats).

Table 3:

Binding behaviour of monoclonal anti-MUC1 antibodies towards the immunodominant PDTR motif: Binding behaviour towards MUC1 peptides glycosylated at the PDTR and non-glycosylated MUC1 peptides of different lengths (1-6 tandem repeats) dependent on the type of immunogen.

Binding behaviour of the antibodies dependent on:

A. Immunogen: MUC1 from tumour material

| | glycosylation | | | | length | | | | length with glyco- sylation |
|----------|---------------|------|-----|----|--------|------|------|----|--------------------------------------|
| | GD-1 | GD-2 | iGD | GI | LD-1 | LD-2 | LD-3 | LI | |
| A76-A/C7 | + | | | | + | | | | + |
| VU-11E2 | + | | | | + | | | | + |
| VU-11D1 | + | | | | + | | | | (+) |
| Ma552 | | + | | | + | | | | - |
| VU-3C6 | | + | | | + | | | | - |
| BC4E549 | | + | | | | + | | | (+) |
| VU-12E1 | | + | | | | + | | | (+) |
| VU-3D1 | | + | | | | | + | | (+) |
| b-12 | | + | | | | | + | | (+) |
| B27.29 | | + | | | | | + | | - |
| MF 06 | | + | | | | | + | | - |

B. Immunogen:

MUC1 peptide (non-glycosylated) or preparations from non-tumour material

| | glycosylation | | | | length | | | | length with glyco- sylation |
|--------|---------------|------|-----|----|--------|------|------|----|--------------------------------------|
| | GD-1 | GD-2 | iGD | GI | LD-1 | LD-2 | LD-3 | LI | |
| VU-4H5 | | | + | | | + | | | + |
| HMPV | | | + | | | + | | | (+) |
| HMFG-1 | | | | + | | + | | | - |
| BC2 | | | | + | | + | | | - |
| VA2 | | | | + | | + | | | - |
| Mc5 | | | | + | | + | | | - |
| E29 | | | | + | | | + | | - |
| BC3 | | | | + | | | + | | - |
| 214D4 | | | | + | | | + | | - |

| | | | | | | | | | |
|---------|--|---|--|---|--|--|---|---|-----|
| BCP8 | | | | + | | | + | | - |
| VA1 | | | | + | | | + | | - |
| C595 | | | | + | | | | + | - |
| BC4W154 | | + | | | | | | + | (+) |

The results show that most antibodies which were produced by an immunisation from mice with tumour material are positively dependent on the increasing length of PDTR-glycosylated MUC1 glycopeptides. This corresponds to an additive effect of the antibodies with regard to the PDTR glycosylation and the length with multiple tandem repeats. A subgroup (A76-A/C7, VU-11E2, VU-11D1) shows a particularly strong additive effect in that the linkage of the antibodies to the MUC1 glycopeptides with several PDTR-glycosylated tandem repeats is clearly increased up to many times over (see Fig. 3A) in comparison with the linkage to short PDTR-glycosylated MUC1 glycopeptides (as in example1) as well as in comparison with the linkage to non-glycosylated MUC1 peptides with several tandem repeats (as in example 2). Antibodies with an additive binding effect exhibit a particularly high affinity towards natural tumour MUC1 consisting of multiple tandem repeats.

As a rule, the antibodies which were produced by an immunisation of mice with non-tumour material and which are PDTR glycosylation-independent show no additive effect, not even when the bonds are dependent on length with regard to non-glycosylated MUC1 peptides (LD-3 and LD-2).

Figure 3B shows an example of an antibody bond without additive effect.

Antibodies VU-4H5 and HMPV which show an iGD linkage show an additive effect described in detail in example 4 as a special case.

Example 3: Binding of MUC1-specific antibodies of the GID group to glycopeptides glycosylated to PDTR with tandem repeats

The method is the same as the one described under example 3.

The results show that the linkage of the VU-4H5 antibody from the GID group depends on the number of the tandem repeats per MUC1 glycopeptide. This result is surprising since this antibody practically does not bind to short DTR-glycosylated

MUC1 peptides. However, it increasingly binds better to DTRP-glycosylated MUC1 glycopeptides with increasing number of tandem repeats. This antibody shows a similar additive effect as the one described in example 3 and, thus, takes an intermediate position, which can also be used for various applications within the meaning of the invention.

Figure 4 shows the linkage of the VU-4H5 antibody to glycosylated MUC1 peptides with different numbers of tandem repeats.

Example 5A) Production of immunostimulatory MUC1 carrying the MUC1 tumour epitope for vaccination

a) Identification of human cells strongly expressing the MUC1 with the tumour epitope

ZR-75-1 cells (ATCC:CRL 1500) were cultivated in RPMI medium with 10% FKS and 1 % glutamine at 37°C, 8% CO₂ and 95% humidity of atmosphere. Compared to other cell lines, these cells already express large amounts of the protein with the tumour epitope on the cell surface. Figure 5A shows immunocytochemical colouring of the ZR-75-1 cells with the A76-A/C7 antibody. The immunocytochemical colouring was carried out by dripping cells on an object carrier, which were then immobilised by incubation in the cell culture incubator in a humid chamber for 30 min and, subsequently, on the object carrier at room temperature for 15 min. The cells were fixed in 5% formaline in PBS for 5 min. After washing the cells with PBS three times, they were incubated with the primary antibody at room temperature for 1.5 hrs, which was diluted in the form of a hybridoma culture supernatant 1:5. After washing the cells three times again with PBS, the secondary antibody (anti-mouse IgG from the rabbit coupled to Cy3, Dianova) was added in a dilution of 1:1000 and incubated for another 30 min at room temperature. After washing the cells three times again, the embedding took place in the Mowiol solution (Calbiochem, Heidelberg, Germany). The isotypic IgG antibody MOPC-21 (Sigma) was used as negative control. The immunocytochemical colouring of the ZR-75-1 cells with the A76-A/C7 made clear that the cell population was not homogenous with regard to the expression of the tumour epitope (Fig. 5A1). In order to increase the expression and to obtain a homogenous A76-A/C7 positive ZR-75-1 cell population, the A76-A/C7 positive ZR-75-1 cells were selected by magnetobeads and, subsequently, clones were cloned by limited dilution, which carry a particularly strong and

homogenous expression of suitable MUC1 molecules. The selection took place by binding of 10^7 cells and the A76-A/C7 antibody (50 μ l of the hybridoma culture supernatant) and anti-IgM antibodies coupled to magnetobeads as well as MS columns from Miltenyi Biotec GmbH (Bergisch Gladbach, Germany) under standard conditions (MACS see product description as to MS columns). The selected cells were cloned by limited dilution in that the cells were adjusted on a cell titre of 150 to 300 cells per 15 ml culture medium. 100 μ l of this dilution is distributed on a microtitre plate in such a way that 1-2 cells per well are cultivated. The distribution on the microtitre plate is checked in the microscope. In this manner, ZR-75-1 sublines could be isolated which express the tumour epitope homogeneously and to a larger extent, as is shown in the immunocytochemical colouring in Figure 5A2.

Figure 5B shows that, in comparison with other cell lines (e.g.: MCF-7), the MUC1 protein, too, is discharged into the cell culture medium in large amounts. This is the result from ELISA analyses with the HMFG-1 (clone 1.10.F3, Beckman/Coulter) as catcher antibody and the A76-A/C7 as detection antibody (Figure 5B). The samples for this ELISA were obtained by sowing 10^5 cells per ml cell culture medium, culturing them under standard conditions for 3 days (see above), subsequently taking an aliquot each day and separating the cell culture supernatant from the cell pellet by centrifugation. 50 μ l of these supernatants were undiluted or used in the dilutions stated (Figure 5B) in the ELISA. The HMFG-1-A76-A/C7 sandwich ELISA was carried out by coating microtitre plates with the catcher antibody HMFG-1, diluted 1:250 in PBS (no indications as to concentration by the manufacturer), in 50 μ l PBS per well at 4°C overnight. Subsequently, the coated plates were blocked twice with 0.1% Tween 20 in PBS at room temperature. The blocking buffer was removed and the plates were washed with washing buffer again three times so as to then put 50 μ l of the samples, undiluted or with the cell culture medium in the dilutions stated, in each well and incubate it at room temperature for 1.5 hrs. The primary antibody was substituted by cell culture medium or 2% BSA, 0.1% Tween 20 in PBS as negative control. After washing it with washing buffer three times, the biotinylated, affinity-purified A76-A/C7 antibody was added in a 1:500 dilution and incubated at room temperature for another hour. After washing it three times again, the binding of streptavidin (1:500 undiluted) coupled to horseradish peroxidase took place at room temperature for 20 min. Finally, the plates were washed in washing buffer twice and in PBS once. The colouring reaction took place in 50 μ l 50 mM acetate buffer pH 6.0 with 1 mg/ml TMB (3,3', 5,5' tetramethylbenzidine, stock solution: 100 mg/ml in DMSO, Sigma) and 0.025% H_2O_2 (Sigma) in the dark at

room temperature for 20 min. The colouring reaction was stopped by adding 25 µl 2.5 N sulfuric acid (final concentration 0.07 N) and measured in the ELISA reader at 450 nm with a reference filter of 620 nm.

The glycosylation of the secreted MUC was analysed in the ELISA by means of different antibodies. In this context, in comparison with the HMFG-1-A76-A/C7 sandwich ELISA described above, the A76-A/C7 antibody was exchanged for antibodies binding the tumour-associated carbohydrates Tn (HB-1, Dako, Glostrup, Denmark) or TF (A78-G/A7, U. Karsten et al, 1995) with high affinity. The detection of the secondary antibody (HB-1 and A78-G/A7) took place with an anti-mouse IgM (diluted 1:5,000) coupled to a POD. The colouring reaction was carried out in 25 mM citric acid, phosphate buffer pH 5.0 with 0.04% H₂O₂ and 0.4 mg/ml o-phenylenediamin (Sigma) in the dark at room temperature for 20 min, stopped by adding 2.5 N sulfuric acid (final concentration 0.07 N) and measured in the ELISA reader at 490 nm with a reference filter of 630 nm. It was possible to detect the tumour-associated carbohydrate epitope Tn and TF, the latter only after treatment with neuraminidase of the MUC1.

b) Overexpression of secretory tumour-associated MUC1 molecules

The overexpression of the secretory MUC1 was achieved by integration of the MUC1 cDNA, whose expression is under the control of a strong promoter (Figure 5C), into a site in the genome of the ZR-75-1 cells which is particularly transcription-active. First, the cells were transfected with the recsite vector which, after integration of the recsite into the genome achieved by antibiotics selection, mediates a stable expression of the LacZ gene. The stably transfected cells were cloned by limited dilution (see next paragraph) and the cells showing the highest LacZ activity were identified. These cells allowed the integration of the MUC1 cDNA with the CMV promoter into a transcription-active site in the genome via the recsite (Figure 5D). The MUC1 cDNA was amplified by means of RT-PCR from ZR-75-1 cells and cloned by restrictions into the expression vector.

Cloning of cells by limited dilution

The cloning of cells by limited dilution is carried out by adjusting the cells on a cell titre of 150 to 300 cells per 15 ml culture medium. 100 µl of this dilution is spread on a microtitre plate in such a way that 1-2 cells per well are cultivated. This distribution on the microtitre plate is checked in the microscope.

c) Purification of the secretory MUC1

The secretory MUC1 was purified from several liters cell culture supernatant by

means of an A76-A/C7 affinity chromatography column under standard conditions. The analysis of the purified protein resulted in several bands in the SDS polyacrylamide gel, which co-migrated with a commercially available MUC1 preparation (Figure 5E).

Example 5B) Influence of the T-cell activation by purified secretory MUC1 carrying the MUC1 tumour epitope

By means of the secreted tumour-associated MUC1 purified with A76-A/C7, incubation was carried out with accumulated T-cells (see above) together with irradiated PBLs. T-cells and PBLs came from two different patients so that an allogene reaction was caused, which was detected by means of T-cell proliferation in BrdU-Assay (see above). In the presence of purified secreted tumour-associated MUC1, no indications were found as to an immunosuppressive effect of MUC1 in this mixed leukocyte reaction. On the contrary, a clear stimulatory effect could be observed at the higher concentration. Reference substances were PBS and 10 µg/ml bovine submaxillary mucine (BSM) respectively (Figure 5F).

Example 6

Production of a monoclonal antibody by means of immunostimulatory MUC1 purified with A76-/C7, which exhibits a particularly strong additive binding effect.

1. Origin of the initial cells

- a) The spleen cells were obtained under sterile conditions (Laminarbox) from a female mouse of the inbred strain Balb/c, which was immunised according to 2.
 - b) X63-Ag8.653 (plasmocytom cell line of Balb/c; Kearney et al., J. Immunol. 123:1548-1550, 1979) was used as fusion cell line. Maintenance took place in medium RPMI 1640 while adding 10% inactivated foetal calf serum and 5×10^{-5} M β -mercaptoethanol.
 - c) Peritoneal cells from Balb/c mice of the same origin as under a) served as feeder cells. They were taken the day before the fusion experiment and/or the cloning under sterile conditions according to a published method and sown into cell culture microtitre plates (Karsten, in: Friemel, Immunologische Arbeitsmethoden, 4th edition, Jena 1991, pp. 309-320).
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2. Immunisation

100 µg MUC1 served as immunogen, which was obtained by means of affinity chromatography with A76-A/C7 (see example 5). MUC1 was applied in PBS with incomplete Freund's adjuvans i.p. and boosted with MUC1 purified with A76-A/C7 5 and 9 weeks later, respectively.

3. Generation of the hybridomas

The hybridoma technique was carried out according to published standard methods (Peters et al., Monoklonale Antikörper, Herstellung und Charakterisierung, Berlin, 1985). The Fusion took place 4 days after the last booster. PEG was used as fusogenic agent. The selection of the hybridomas took place by means of azaserine/hypoxanthine; mouse peritoneal cells were used for support of growth. The forming hybridomas were tested by means of ELISA with immobilised MUC1, MUC1 glycopeptides and in immunofluorescence tests with MUC1 positive cell lines (for example T-47D-cells, ZR-75-1 cells and subcell lines) on the secretion of specific antibodies.

Cloning took place by means of limiting dilution according to standard techniques in the presence of mouse peritoneal cells (Karsten, in: Friemel, Immunologische Arbeitsmethoden, 4th edition, Jena 1991, pp. 309-320). The isotypes were detected by means of a commercial isotypes-ELISA-kit (Pharmigen).

Example 7: Tests as to the tumour-specific cytotoxicity

a) Identification and generation of a subpopulation of human MCF-7 cells which strongly express MUC1 with the tumour epitope

MCF-7 cells (ATCC:HTB-22) were - as described in detail under example 5 for ZR-75-1 - cultivated and isolated from the cells by means of magnetobeads with A76-A/C7 cells and, subsequently, clones carrying a particularly strong and homogenous expression of suitable MUC1 molecules were cloned by limited dilution. In this way, MCF-7 sublines could be isolated, which expressed the tumour epitope in a homogenously and stronger than the initial line.

b) Generation of mature Nemod-mDC cells charged with lysates from MCF-7

As to dendritic cells, effective dendritic cells (DE 10139428.4) from NemodDC-1 cells stemming from MUTZ-3 were produced. Effective dendritic cells were

differentiated by differentiation from NemodDC-1 cells (precursor cells, prec-NemodDC) to immature dendritic cells (i-NemodDC), charged with antigen and, subsequently, matured to mature effective dendritic cells (m-NemodDC) which are able to activate T-cells in an antigen-specific way. I-NemodDC which are charged with antigens absorb these cells, process them and present peptide fragments of the antigens in the context of their MHC class molecules, for example MHC I-A2. By means of these effective dendritic cells, both specific CTL- and helper T-cells from naïve T-cells can be stimulated and NKT-cells can be stimulated and expanded.

4×10^6 prec-Nemod-DC cells were differentiated in 40 ml reaction volume in α MEM + 20% FCS + 10% CM5637 with 1,000 U/ml GM-CSF, 1,000 U/ml IL-4 and 2.5 ng/ml TNF α for 7 days. Subsequently, the cells (I-NemodDC) were charged with a suitable amount of tumour cell lysate from the MCF-7-cells described above - which were produced by alternating freezing and thawing by means of liquid nitrogen - for 6 hrs and, subsequently, matured with TNF α for another 24 hrs, irradiated (30 Gy) and used for stimulation of T-lymphocytes.

c) Isolation of CD8+-T-cells by means of microbeads

CD8+-T-cells cells HLA-A2 from positive donors were purified via a MAC isolation by means of magnetbeads coated with antibodies (Miltenyi, manufacturer's instructions) according to known methods from PBMC from usual buffy coat preparations.

d) Generation of cytotoxic T-cells (CTLs)

The CD8+-T-lymphocytes (1×10^6 CD8+-T-lymphocytes/ml) of a HLA-A2 positive donor were stimulated weekly with m-NemodDC (ratio 1:10) charged with tumour cell lysate (MCF7 lysates) in medium (AIM V + 10 U/ml IL-2 from day 7) (prime + restimulation).

e) Cytotoxicity test

First, the target cells(MCF-7, LS 174 T and T2) which are in a physiologically stable and vital state are washed in cold medium (DMEM + 5% FCS) and, after centrifugation (300 x g, 7 min, RT) added to europium buffer (50 mM HEPES, 93 mM sodium chloride, 5 mM potassium chloride, 2 mM magnesium chloride, 10 mM DTPA, 2 mM europium-III-acetate). During this process, 5×10^6 target cells with

a volume of 800 µl were transferred into an electroporation cuvette (d = 4 mm). Before and after the electroporation, an incubation period of 6 to 10 min on ice took place. The electroporation was carried out with an Eppendorf Multiporator (30 µs, 710 V or 1,100 V and 1 puls). After washing the cells with medium five times, the marked target cell is ready for use in the cytotoxicity test. For this purpose, it was either used directly after the determination of the vitality and cell density (MCF-7 or LS 174 T) or marked with peptide (MUC1 A2 peptide: e.g. LLLLTVLTL; irrelevant HIV-gag A2 peptide; 5 µg/ml each) (T2 cells) for another 2 hours so as to analyse peptide-specific effector cells. For the cytotoxicity test, marked target cells (5 to 10 x 10³) were presented in a volume of 100 µl/well in a microtitre plate with an arch. Subsequently, effector cells, also in a volume of 100 µl, were added at different cell concentrations so that the result of the ratio of effector cell and target cell was 1.25:1 to 100:1. Apart from these solutions, samples were prepared for the determination of the background signal (BR) of the spontaneous and maximum release. The background signal was a cell-free medium sample at the beginning of the experiment. The spontaneous release (SR) allows assessment of the stability of the marking of the target cell with europium. For the determination, marked target cells were cultivated without effector cells and, after the end of the incubation period, the concentration of the released europium was determined. The maximal release (MR) was determined by means of marked target cells which were incubated, also without effector cells but with an artificial reagent (here with 50% of the purest ethanol), for the testing period and brought to total lysis. According to the amounts defined above, the percentage of the spontaneous release was calculated, which, for the assessment of a successful test, should be below 30%. Furthermore, the percentage of the specific, cytotoxic activity of the effector cell was characterised.

$$\% \text{ spontaneous release} = \frac{SR \text{ (counts)} - BR \text{ (counts)}}{MR \text{ (counts)} - BR \text{ (counts)}} \times 100$$

$$\% \text{ specific release} = \frac{\text{effector /target cell (counts)} - SR \text{ (counts)}}{MR \text{ (counts)} - SR \text{ (counts)}} \times 100$$

After the incubation period of 4 hours, the culture supernatant containing the

europium was separated by means of centrifugation of the microtitre plate (500 x g, 5 min, RT). Subsequently, 20 µl of this culture supernatant were transferred to a presented microtitre plate with a flat bottom with 200 µl enhancement solution/well and incubated on a plate shaker for another 15 min. During this incubation period, a fluorescent complex results from the europium DTPA chelate under formation of a europium-NTA chelate. The samples were measured in a time-delayed fluorometer (Victor², PerkinElmar) at a wave length of 613 nm.

In the experiment, MCF-7 was used as relevant target cell (Figure 6, MCF7) and LS 174 T (Figure 6, LS 174T) was used as irrelevant target cell. Furthermore, T2 cells which were not able to process and which can only be charged on their MHC class I A2 molecules by defined A2 peptides were used. In this context, a MUC1-A2-peptide was used for testing an effective stimulation of originally naïve T-cells which are specifically directed against MUC1 by means of the MUC1 molecules of the invention, which, in the present example, are present in the corresponding cell lysates (T2 + MUC1). An irrelevant HIVgag A2 peptide was used as negative control (T2 + HIV).

The data show that by means of the MUC1 molecules of the invention in a cell lysate a MUC1-specific cytotoxic immune response can be achieved against cells carrying MUC1 on the surface in the context of HLA-A2. These immunological data show that the MUC1 molecules of the invention can be used both as such for the activation of an effective anti-tumour response as well as in form of dendritic cells which were charged with the MUC1 molecules of the invention. Moreover, the data show that MUC1-specific T-cells can be ex vivo expanded and specifically activated by means of the MUC1 molecules and dendritic cells of the invention, which demonstrates a suitability for adoptive T-cell therapies.
